

зинового взаимодействия в присутствии мутаций в мышечных белках, связанных с врожденными миопатиями, что показано нами на примере тропомиозина с заменой R90P, встроеного в мышечное волокно. Патологическое действие мутации R90P связано с увеличением АТФазной активности миозина и относительного количества головок миозина, которые находятся в конформации сильного связывания, при моделировании различных стадий цикла гидролиза АТФ (Borovikov et al., 2021). Одним из главных эффектов мутации являлось снижение амплитуды движения головок миозина (или SH1-спирали миозина) в цикле гидролиза АТФ. Добавление ВДМ к мышечным волокнам, содержащим мутантный тропомиозин, частично нормализовало конформационные перестройки миозина, нарушенные в присутствии мутантного тропомиозина, позволяя головкам миозина эффективнее переходить между конформациями слабого и сильного связывания с актином. Из данных следует, что ВДМ может быть использован для восстановления нормальной регуляции взаимодействия миозина с актином и должен быть протестирован в дальнейшем на других модельных системах и в модельных животных.

#### ФИНАНСИРОВАНИЕ РАБОТЫ

Работа выполнена при финансовой поддержке РФФИ (проект № 20-04-00523).

#### СОБЛЮДЕНИЕ ЭТИЧЕСКИХ СТАНДАРТОВ

Все процедуры с лабораторными животными были проведены по правилам, одобренным Комиссией по биоэтике Института цитологии РАН (№ F18-00380, 12.10.2017–31.10.2022).

#### КОНФЛИКТ ИНТЕРЕСОВ

Авторы заявляют об отсутствии конфликта интересов.

#### СПИСОК ЛИТЕРАТУРЫ

- Andreev O.A., Takashi R., Borejdo J.* 1995. Fluorescence polarization study of the rigor complexes formed at different degrees of saturation of actin filaments with myosin subfragment-1. *J. Muscle Res. Cell Motil.* V. 16. P. 353.
- Borejdo J., Assulin O., Ando T., Putnam S.* 1982. Cross-bridge orientation in skeletal muscle measured by linear dichroism of an extrinsic chromophore. *J. Mol. Biol.* V. 158. P. 391. [https://doi.org/10.1016/0022-2836\(82\)90205-4](https://doi.org/10.1016/0022-2836(82)90205-4)
- Borovikov Y.S., Andreeva D.D., Avrova S.V., Sirenko V.V., Simonyan A.O., Redwood C.S., Karpicheva O.E.* 2021. Molecular mechanisms of the deregulation of muscle contraction induced by the R90P mutation in Tpm3.12 and the weakening of this effect by BDM and W7. *Int. J. Mol. Sci.* V. 22. P. 6318. <https://doi.org/10.3390/ijms22126318>
- Borovikov Y.S., Dedova I.V., dos Remedios C.G., Vikhoreva N.N., Vikhorev P.G., Avrova S.V., Hazlett T.L., Van Der Meer B.W.* 2004. Fluorescence depolarization of actin filaments in re-constructed myofibers: the effect of S1 or pPDM-S1 on movements of distinct areas of actin. *Biophys. J.* V. 86. P. 3020. [https://doi.org/10.1016/S0006-3495\(04\)74351-9](https://doi.org/10.1016/S0006-3495(04)74351-9)
- Borovikov Y.S., Karpicheva O.E., Avrova S.V., Redwood C.S.* 2009. Modulation of the effects of tropomyosin on actin and myosin conformational changes by troponin and Ca<sup>2+</sup>. *Biochim. Biophys. Acta.* V. 1794. P. 985. <https://doi.org/10.1016/j.bbapap.2008.11.014>
- Borovikov Y.S., Rysev N.A., Avrova S.V., Karpicheva O.E., Borys D., Moraczewska J.* 2017. Molecular mechanisms of deregulation of the thin filament associated with the R167H and K168E substitutions in tropomyosin Tpm 1.1. *Arch. Biochem. Biophys.* V. 614. P. 28. <https://doi.org/10.1016/j.abb.2016.12.004>
- Bradford M.M.* 1976. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.* V. 72. P. 248. <https://doi.org/10.1006/abio.1976.9999>
- Burghardt T.P., Garamszegi S.P., Ajtai K.* 1997. Probes bound to myosin Cys-707 rotate during length transients in contraction. *Proc. Natl. Acad. Sci. USA.* V. 94. P. 9631. <https://doi.org/10.1073/pnas.94.18.9631>
- Fiske C.H., Subbarow Y.* 1925. Determination of inorganic phosphate. *J. Biol. Chem.* V. 66. P. 375.
- Herrmann C., Wray J., Travers F., Barman T.* 1992. Effect of 2,3-butanedione monoxime on myosin and myofibrillar ATPases. An example of an uncompetitive inhibitor. *Biochemistry.* V. 31. P. 12227. <https://doi.org/10.1021/bi00163a036>
- Higuchi H., Takemori S.* 1989. Butanedione monoxime suppresses contraction and ATPase activity of rabbit skeletal muscle. *J. Biochem.* V. 105. P. 638. <https://doi.org/10.1093/oxfordjournals.jbchem.a122717>
- Karpicheva O.E., Sirenko V.V., Rysev N.A., Simonyan A.O., Borys D., Moraczewska J., Borovikov Y.S.* 2017. Deviations in conformational rearrangements of thin filaments and myosin caused by the Ala155Thr substitution in hydrophobic core of tropomyosin. *Biochim. Biophys. Acta.* V. 1. P. 1790. <https://doi.org/10.1016/j.bbapap.2017.09.008>
- Komatsu H., Koseki Y., Kanno T., Aoki S., Kodama T.* 2017. 2,3-Butandione 2-monoxime inhibits skeletal myosin II by accelerating ATP cleavage. *Biochem. Biophys. Res. Commun.* V. 490. P. 849. <https://doi.org/10.1016/j.bbrc.2017.06.130>
- Lawlor M.W., Dechene E.T., Roumm E., Geggel A.S., Moghadaszadeh B., Beggs A.H.* 2010. Mutations of tropomyosin 3 (TPM3) are common and associated with type 1 myofiber hypotrophy in congenital fiber type disproportion. *Hum. Mutat.* V. 31. P. 176. <https://doi.org/10.1002/humu.21157>
- Lee B.K., Jeung K.W., Choi S.S., Park S.W., Yun S.W., Lee S.M., Kim N.Y., Heo T., Min Y.I.* 2015. Effects of the administration of 2,3-butanedione monoxime during conventional cardiopulmonary resuscitation on ischaemic contracture and resuscitability in a pig model of out-of-hospital cardiac arrest. *Resuscitation.* V. 87. P. 26. <https://doi.org/10.1016/j.resuscitation.2014.11.011>

- Margossian S., Lowey S.* 1982. Preparation of myosin and its subfragments from rabbit skeletal muscle. *Methods Enzym.* V. 85. P. 55.
- McKillop D.F., Fortune N.S., Ranatunga K.W., Geeves M.A.* 1994. The influence of 2,3-butanedione 2-monoxime (BDM) on the interaction between actin and myosin in solution and in skinned muscle fibres. *J. Muscle Res. Cell Motil.* V. 15. P. 309.  
<https://doi.org/10.1007/BF00123483>
- Nesmelov Y.E., Agafonov R.V., Burr A.R., Weber R.T., Thomas D.D.* 2008. Structure and dynamics of the force-generating domain of myosin probed by multifrequency electron paramagnetic resonance. *Biophys. J.* V. 95. P. 247.  
<https://doi.org/10.1529/biophysj.107.124305>
- Okamoto Y., Sekine T.* 1985. A streamlined method of subfragment one preparation from myosin. *J. Biol. Chem.* V. 98. P. 1143.
- Potter J.D.* 1982. Preparation of troponin and its subunits. *Methods Enzym.* V. 85. P. 241.
- Reedy M.K., Holmes K.C., Tregear R.T.* 1965. Induced changes in orientation of the cross-bridges of glycerinated insect flight muscle. *Nature.* V. 207. P. 1276.  
<https://doi.org/10.1038/2071276a0>
- Robinson P., Lipscomb S., Preston L.C., Altin E., Watkins H., Ashley C.C., Redwood C.S.* 2007. Mutations in fast skeletal troponin I, troponin T, and beta-tropomyosin that cause distal arthrogryposis all increase contractile function. *FASEB J.* V. 21. P. 896.  
<https://doi.org/10.1096/fj.06-6899com>
- Roopnarine O., Thomas D.D.* 1996. Orientation of intermediate nucleotide states of indane dione spin-labeled myosin heads in muscle fibers. *Biophys. J.* V. 70. P. 2795.
- Spudich J.A., Watt S.* 1971. The regulation of rabbit skeletal muscle contraction. I. Biochemical studies of the interaction of the tropomyosin-troponin complex with actin and the proteolytic fragments of myosin. *J. Biol. Chem.* V. 246. P. 4866.
- Takezawa Y., Kim D.S., Ogino M., Sugimoto Y., Kobayashi T., Arata T., Wakabayashi K.* 1999. Backward movements of cross-bridges by application of stretch and by binding of MgADP to skeletal muscle fibers in the rigor state as studied by X-ray diffraction. *Biophys. J.* V. 76. P. 1770.  
[https://doi.org/10.1016/S0006-3495\(99\)77338-8](https://doi.org/10.1016/S0006-3495(99)77338-8)
- Tregear R.T., Mendelson R.A.* 1975. Polarization from a helix of fluorophores and its relation to that obtained from muscle. *Biophys. J.* V. 15. P. 455.
- Volkman N., Hanein D.* 2000. Actomyosin: law and order in motility. *Curr. Opin. Cell Biol.* V. 12. P. 26.  
[https://doi.org/10.1016/s0955-0674\(99\)00053-8](https://doi.org/10.1016/s0955-0674(99)00053-8)
- Wheeler T.J., Chien S.* 2012. Protection of rat cardiac myocytes by fructose-1,6-bisphosphate and 2,3-butanedione. *PLoS One.* V. 7. P. e35023.  
<https://doi.org/10.1371/journal.pone.0035023>

## Influence of 2,3-Butanedione-Monoxime on the Interaction of Myosin with Actin in Healthy and in Congenital Myopathy

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Congenital myopathies are a heterogeneous group of human skeletal muscle disorders characterized by muscle hypotonia and weakness. Myopathies have a wide range of clinical phenotypes, which makes it extremely difficult to develop approaches to their treatment. There are several pharmacological agents in clinical use or under clinical investigation for the treatment of cardiomyopathies whose mechanism of action can be used to treat congenital myopathies as well. One such agent is 2,3-butanedione-monoxime (BDM), a noncompetitive inhibitor of myosin ATPase activity used to suppress acute myocardial injury. The molecular mechanisms of inhibition of myosin by BDM in skeletal muscle have not been studied, therefore the aim of this work was to estimate the effect of BDM on the interaction of myosin with actin in the modeling of several ATPase stages in skeletal muscle fiber, in order to assess the prospects for the use of BDM for the treatment of congenital myopathies. We found that BDM enhances the rigidity of myosin binding to actin when modeling weak binding forms of these muscle proteins, which can slow down the transition of actomyosin from the AM · ADP · Pi to the AM · ADP state and is one of the reasons for the decrease in myosin ATPase activity in the presence of BDM. When modeling successive stages of the ATPase cycle using ADP, AMPPNP, ATPγS, and ATP, the myosin heads gradually switch to a state of weak interaction with actin. In the presence of the regulatory proteins tropomyosin and troponin in the muscle fiber, BDM does not affect the formation of a weak form of actomyosin binding, but increases the number of myosin heads essential for force generation. BDM can be used to increase the efficiency of myosin conformational rearrangements in the presence of tropomyosin with the *R90P* mutation associated with congenital myopathy, since this reagent increases the number of myosin heads in the muscle fiber capable of effective conformational rearrangements in the ATPase cycle and partially inhibits the pathological effects of the mutation.

**Keywords:** actin-myosin interaction, regulation of muscle contraction, muscle fiber, polarized fluorescence, inhibitor of myosin ATPase activity, 2,3-butanedione-monoxime, congenital myopathies