

- tramer is formed at the CD95 DISC. *Cell Death Differ.* V. 10. P. 144.
- Medema J.P., Scaffidi C., Kischkel F.C., Shevchenko A., Mann M., Krammer P.H., Peter M.E., 1997. FLICE is activated by association with the CD95 death-inducing signaling complex (DISC). *EMBO J.* V. 16. P. 2794.
- Muzio M., Chinnaiyan A.M., Kischkel F.C., O'Rourke K., Shevchenko A., Ni J., Scaffidi C., Bretz J. D., Zhang M., Gentz R., Mann M., Krammer P.H., Peter M.E., Dixit V.M. 1996. FLICE, a novel FADD-homologous ICE/CED-3-like protease, is recruited to the CD95 (Fas/APO-1) death-inducing signaling complex. *Cell.* V. 85, P. 817.
- Neumann L., Pforr C., Beaudouin J., Pappa A., Fricker N., Krammer P.H., Lavrik I.N., Eils R. 2010. Dynamics within the CD95 death-inducing signaling complex decide life and death of cells. *Mol. Syst. Biol.* V. 6. P. 352.
- Roberts A.W., Davids M.S., Pagel J.M., Kahl B.S., Puvvada S.D., Gerecitano J.F., Kipps T.J., Anderson M.A., Brown J.R., Gressick L., Wong S., Dunbar M., Zhu M., Desai M.B., Cerri et al. 2016. Targeting BCL2 with venetoclax in relapsed chronic lymphocytic leukemia. *New Eng. J. Med.* V. 374. P. 311.
- Schleich K., Warnken U., Fricker N., Ozturk S., Richter P., Kammerer K., Schnolzer M., Krammer P.H., Lavrik I.N. 2012. Stoichiometry of the CD95 death-inducing signaling complex: Experimental and modeling evidence for a death effector domain chain model. *Mol. Cell.* V. 47. P. 306.
- Seyrek K., Ivanisenko N.V., Richter M., Hillert L.K., Konig C., Lavrik I.N. 2020. Controlling cell death through post-translational modifications of DED proteins. *Trends Cell Biol.* V. 30. P. 354.
- Shen C., Yue H., Pei J., Guo X., Wang T., Quan J.M. 2015. Crystal structure of the death effector domains of caspase-8. *Biochem. Biophys. Res. Commun.* V. 463. P. 297.
- Sprick M.R., Rieser E., Stahl H., Grosse-Wilde A., Weigand M.A., Walczak H. 2002. Caspase-10 is recruited to and activated at the native TRAIL and CD95 death-inducing signalling complexes in a FADD-dependent manner but can not functionally substitute caspase-8. *EMBO J.* V. 21. P. 4520.
- Sprick M.R., Weigand M.A., Rieser E., Rauch C.T., Juo P., Blenis J., Krammer P.H., Walczak H. 2000. FADD/MORT1 and caspase-8 are recruited to TRAIL receptors 1 and 2 and are essential for apoptosis mediated by TRAIL receptor 2. *Immunity.* V. 12. P. 599.
- Sterling T., Irwin J.J. 2015. ZINC 15—Ligand Discovery for Everyone. *J. Chem. Inf. Model.* V. 55. P. 2324.
- Wang L., Yang J.K., Kabaleswaran V., Rice A.J., Cruz A.C., Park A.Y., Yin Q., Damko E., Jang S.B., Raunser S., Robinson C.V., Siegel R.M., Walz T., Wu H. 2010. The Fas-FADD death domain complex structure reveals the basis of DISC assembly and disease mutations. *Nat. Struct. Mol. Biol.* V. 17. P. 1324.
- Wilson N.S., Dixit V., Ashkenazi A. 2009. Death receptor signal transducers: nodes of coordination in immune signaling networks. *Nat. Immunol.* V. 10. P. 348.
- Yu J.W., Jeffrey P.D., Shi Y. 2009. Mechanism of procaspase-8 activation by c-FLIPL. *Proc. Natl. Acad. Sci. USA.* V. 106. P. 8169.

Development of Small Molecules Targeting Procaspase-8 at the DISC

J. Espe^a, N. V. Ivanisenko^b, L. K. Hillert-Richter^a, V. A. Ivanisenko^b and , and I. N. Lavrik^{a, b, *}

^a*Translational Inflammation Research, Medical Faculty, Center of Dynamic Systems, Otto von Guericke University Magdeburg, Magdeburg, 39120 Germany*

^b*Institute of Cytology and Genetics SB RAS, Novosibirsk, 630090 Russia*

**e-mail: inna.lavrik@med.ovgu.de*

Pharmacological targeting *via* small molecule-based chemical probes has recently acquired attention as a valuable tool to dissect molecular mechanisms. Induction of extrinsic apoptosis *via* CD95/Fas and TRAIL-R1/2 is initiated by the formation of the Death-Inducing Signaling Complex (DISC). Procaspase-8 activation at the DISC is a central event triggering extrinsic apoptosis. Procaspase-8 activation takes place at the death effector domain (DED) filaments that are formed at the DISC. Hence, targeting the DED filaments is essential to gain insight into DISC control and pharmacological targeting of extrinsic apoptosis. In this study we developed the group of chemical probes that were designed *in silico* to target procaspase-8 at the DED filaments. This was followed by their experimental validation *via* cell viability assays and selection of the most effective compounds. Taken together, our study describes a development of new chemical compounds that are constructed to target procaspase-8 and extrinsic apoptosis.

Keywords: procaspase-8, small molecules, DED, DISC, apoptosis, caspase activation, DR