- Mandal R., Raab M., Matthess Y., Becker S., Knecht R., Strebhardt K. 2014. pERK 1/2 inhibit caspase-8 induced apoptosis in cancer cells by phosphorylating it in a cell cycle specific manner. Mol. Oncol. V. 8. P. 232.
- *Mebratu Y., Tesfaigzi Y.* 2009. How ERK1/2 activation controls cell proliferation and cell death: Is subcellular localization the answer? Cell Cycle Georget. Tex. V. 8. P. 1168.
- Meyer S., Fuchs T.J., Bosserhoff A.K., Hofstädter F., Pauer A., Roth V., Buhmann J.M., Moll I., Anagnostou N., Brandner J.M., Ikenberg K., Moch H., Landthaler M., Vogt T., Wild P.J. 2012. A Seven-marker signature and clinical outcome in malignant melanoma: A large-scale tissue-microarray study with two independent patient cohorts. PLoS One. V. 7.
- Morin P.J., Vogelstein B., Kinzler K.W. 1996. Apoptosis and APC in colorectal tumorigenesis. Proc. Natl. Acad. Sci. USA. V. 93. P. 7950.
- *Nagata Y., Todokoro K.* 1999. Requirement of activation of JNK and p38 for environmental stress-induced erythroid differentiation and apoptosis and of inhibition of ERK for apoptosis. Blood. V. 94. P. 853.
- *Nejak-Bowen K., Kikuchi A., Monga S.P.S.* 2013. Beta-catenin-NF-κB interactions in murine hepatocytes: A complex to die for. Hepatol. Baltim. Md. V. 57. P. 763.
- Park J.S., Carter S., Reardon D.B., Schmidt-Ullrich R., Dent P., Fisher P.B. 1999. Roles for basal and stimulated p21(Cip-1/WAF1/MDA6) expression and mitogen-activated protein kinase signaling in radiation-induced cell cycle checkpoint control in carcinoma cells. Mol. Biol. Cell. V. 10. P. 4231.
- Pospelova T.V., Medvedev A.V., Kukushkin A.N., Svetlikova S.B., van der Eb A.J., Dorsman J.C., Pospelov V.A. 1999. E1A + cHa-ras transformed rat embryo fibroblast cells are characterized by high and constitutive DNA binding activities of AP-1 dimers with significantly altered composition. Gene Expr. V. 8. P. 19.
- Rasola A., Sciacovelli M., Chiara F., Pantic B., Brusilow W.S., Bernardi P. 2010. Activation of mitochondrial ERK protects cancer cells from death through inhibition of the per-

meability transition. Proc. Natl. Acad. Sci. USA. V. 107. P. 726.

- Shao N., Zou J., Li J., Chen F., Dai J., Qu X., Sun X., Ma D., Ji C. 2012. Hyper-activation of WNT/β-catenin signaling pathway mediates anti-tumor effects of histone deacetylase inhibitors in acute T lymphoblastic leukemia. Leuk. Lymphoma. V. 53. P. 1769.
- Stang S.L., Lopez-Campistrous A., Song X., Dower N.A., Blumberg P.M., Wender P.A., Stone J.C. 2009. A proapoptotic signaling pathway involving RasGRP, Erk, and Bim in B cells. Exp. Hematol. V. 37. P. 122.
- Tammina S.K., Mandal B.K., Ranjan S., Dasgupta N. 2017. Cytotoxicity study of Piper nigrum seed mediated synthesized SnO₂ nanoparticles towards colorectal (HCT116) and lung cancer (A549) cell lines. J. Photochem. Photobiol. B. V. 166. P. 158.
- Wang H., Chi C.-H., Zhang Y., Shi B., Jia R., Wang B.-J. 2019. Effects of histone deacetylase inhibitors on ATP-binding cassette transporters in lung cancer A549 and colorectal cancer HCT116 cells. Oncol. Lett. V. 18. P. 63.
- Wang Y., Tang H., He G., Shi Y., Kang X., Lyu J., Zhou M., Zhu M., Zhang J., Tang K. 2018. High Concentration of aspirin induces apoptosis in rat tendon stem cells via inhibition of the Wnt/β-catenin pathway. Cell. Physiol. Biochem. Int. J. Exp. Cell. Physiol. Biochem. Pharmacol. V. 50. P. 2046.
- *Willert K., Jones K.A.* 2006. Wnt signaling: is the party in the nucleus? Genes Dev. V. 20. P. 1394.
- Yu X.-D., Wang S.-Y., Chen G.A., Hou C.-M., Zhao M., Hong J.A., Nguyen D.M., Schrump D.S. 2007. Apoptosis induced by depsipeptide FK228 coincides with inhibition of survival signaling in lung cancer cells. Cancer J. Sudbury Mass. V. 13. P. 105.
- Zimmerman Z.F., Kulikauskas R.M., Bomsztyk K., Moon R.T., Chien A.J. 2013. Activation of Wnt/β-catenin signaling increases apoptosis in melanoma cells treated with trail. PloS One. V. 8. P. e69593. https://doi.org/10.1371/journal.pone.0069593

THE INFLUENCE OF A SODIUM BUTYRATE ON PROLIFERATIVE SIGNALING CASCADES IN SENSITIVE AND RESISTANT TO HDAC INHIBITORS ACTION CELLS

O. O. Gnedina^{*a*} and M. V. Igotti^{*a*}, *

^aInstitute of Cytology of the Russian Academy of Sciences, St. Petersburg, 194064 Russia

*e-mail: marie.igotti@gmail.com

To establish the mechanisms of transformed cells resistance to the histone deacetylase inhibitors (HDACi), we compared the changes of the main proliferative signaling cascades activities in cells that are sensitive or resistant to HDA-Ci-induced apoptosis. The time-dependent dynamics of the ERK kinase activity was shown. Phosphorylation of ERK kinase increased in the first 24 hours of the HDACi sodium butyrate treatment, followed by ERK activity decrease in resistant cells. Whereas in apoptotic cells, an inverse time-dependent dynamics of ERK activity changes was observed. It has been shown that resistance to HDACi can be overcome by inhibiting the MEK/ERK pathway. The resistant cells underwent to apoptotic death after 48 hours of combined treatment with sodium butyrate and the MEK/ERK pathway inhibitor PD098059. The study of the Wnt/ β -catenin signaling cascade showed that the accumulation and transcriptional activation of β -catenin occurs only in cells resistant to HDACi-induced apoptosis. Thus, the obtained results indicate that a change in the activity of β -catenin is one of the reasons for the resistance to apoptosis induced by HDACi sodium butyrate, and the increased activity of the PI3K/Akt and MEK/ERK kinase pathways is a prerequisite for the most effective antiproliferative effect of HDACi.

Keywords: tumor cells, histone deacetylase inhibitor (HDACi), apoptosis, resistance, β -catenin, PKB/Akt and ERK kinases

ЦИТОЛОГИЯ том 62 № 11 2020