chronic renal failure in the Russian Federation in 1998–2007 (Analytical report according to the Russian Register of Renal Replacement Therapy). Nephrol. dialysis. V. 13. N° 3. P. 150.)

- Acland R.D. 1972. Signs of patency in small vessel anastomosis. Surgery. V. 72. P. 744–748.
- Drews J.D., Miyachi H., Shinoka T. 2017. Tissue-engineered vascular grafts for congenital cardiac disease: Clinical experience and current status. Trends Cardiovasc. Med. V. 27 P. 521–531.
- Fukunishi T., Best C.A., Sugiura T., Opfermann J., Ong C.S., Shinoka T., Breuer C.K., Krieger A., Johnson J., Hibino N. 2017. Preclinical study of patient-specific cell-free nanofiber tissue-engineered vascular grafts using 3-dimensional printing in a sheep model. J. Thorac. Cardiovasc. Surg. V. 153. P. 924–932.
- Harskamp R.E., Lopes R.D., Baisden C.E., De Winter R.J., Alexander J.H. 2013. Saphenous vein graft failure after coronary artery bypass surgery: pathophysiology, management, and future directions. Annals Surgery. V. 257. P. 824–833.
- Klinkert P., Post P.N., Breslau P.J., van Bockel J.H. 2004. Saphenous vein versus PTFE for above-knee femoropopliteal bypass. A review of the literature. Eur. J. Vasc. Endovasc. Surg. V. 27. P. 357–362.
- Konig G., McAllister T.N., Dusserre N., Garrido S.A., Iyican C., Marini A., Fiorillo A., Avila H., Wystrychowski W., Zagalski K., Maruszewski M., Jones A.L., Cierpka L., de la Fuente L.M., L'Heureux N. 2009. Mechanical properties of completely autologous human tissue engineered blood vessels compared to human saphenous vein and mammary artery. Biomaterials. V. 30. P. 1542–1550.
- L'Heureux N., Dusserre N., Konig G., Victor B., Keire P., Wight T.N., Chronos N.A., Kyles A.E., Gregory C.R., Hoyt G., Robbins R.C., McAllister T.N. 2006. Human tissue-engineered blood vessels for adult arterial revascularization. Nat. Med. V. 12. P. 361–365.
- Norotte C., Marga F.S., Niklason L.E., Forgacs G. 2009. Scaffold-free vascular tissue engineering using bioprinting. Biomaterials. V. 11. P. 35–38.

- Popryadukhin P.V., Popov G.I., Yukina G.Yu., Dobrovolskaya I.P., Ivan'kova E.M., Vavilov V.N. Yudin V.E. 2017. Tissue-engineered vascular graft of small diameter based on electrospun polylactide microfibers. Int. J. Biomater. Article ID 9034186. https://doi.org/10.1155/2017/9034186
- Row S., Santandreu A., Swartz D.D., Andreadis S.T. 2017. Cellfree vascular grafts: Recent developments and clinical potential. Technol. (Singap. World Sci.). V. 5. P. 13–20.
- Shinoka T., Breuer C. 2008. Tissue-engineered blood vessels in pediatric cardiac surgery. Yale J. Biol. Med. V. 81. P. 161–166.
- Smith R.J.Jr., Yi T., Nasiri B., Breuer C.K., Andreadis S.T. 2019. Implantation of VEGF-functionalized cell-free vascular grafts: regenerative and immunological response. FASEB J. V. 33. P. 5089–5100.
- Sullivan S.J., Brockbank K.G.M. 2000. Small-diameter vascular grafts. In: Principles of tissue engineering. N.-Y.: Academic Press. P. 447–453.
- Tara S., Kurobe H., Rocco K.A., Maxfield M.W., Best C.A., Yi T., Naito Y., Breuer C.K., Shinoka T. 2014. Well-organized neointima of large-pore poly(L-lactic acid) vascular graft coated with poly(L-lactic-co-ε-caprolactone) prevents calcific deposition compared to small-pore electrospun poly(L-lactic acid) graft in a mouse aortic implantation model. Atherosclerosis. V. 237. P. 684–691.
- Vaz C.M., van Tuijl S., Bouten C.V., Baaijens F.P. 2005. Design of scaffolds for blood vessel tissue engineering using a multi-layering electrospinning technique. Acta Biomater. V. 1. P. 575–582.
- Watanabe T., Kanda K., Ishibashi-Ueda H., Yaku H., Nakayama Y. 2010. Autologous small-caliber "biotube" vascular grafts with argatroban loading: A histomorphological examination after implantation to rabbits. J. Biomed. Mater. Res. B Appl. Biomater. V.92. P. 236–242.
- Zhao Y., Zhang S., Zhou J., Wang J., Zhen M., Liu Y., Chen J., Qi Z. 2010. The development of a tissue-engineered artery using decellularized scaffold and autologous ovine mesenchymal stem cells. Biomaterials. V. 31. P. 296–307.

MORPHOLOGICAL STUDY OF A BIORESORBABLE TUBULAR MATRIX OF A SMALL DIAMETER FROM A POLY(L-LACTIDE) FOR A TISSUE-ENGINEERED VASCULAR IMPLANT

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Non-woven tubular bioresorbable matrices with an inner diameter of 1.1 mm were obtained by electrospinning from solutions of poly(L-lactide) (PLA). Matrices were implanted in the abdominal part of the aorta to rats as a tissue-engineered vascular implant for a period of 2 days to 16 months and showed high biocompatibility, non-toxicity, and pronounced atrombogenic properties. The total implant patency was 93%. Morphometric analysis of the dynamics of population of the matrix with cells showed that at all periods of observation in the outer half of the matrix wall the number of cells prevails, which indicates the migration of cells from the connective tissue capsule surrounding the

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matrix. It was shown that two parallel processes occur in the matrix wall: bioresorption of PLA fibers and the formation of connective tissue. Complete bioresorption of matrices with the replacement of native tissues, the formation of the endothelial and subendothelial layers took place over the 16 months of the experiment. By this time, all experimental animals in the reconstruction zone had aneurysmal enlargement that did not lead to rupture of the implant. In order to prevent the development of such complications, it is necessary to develop a method for additional strengthening of the matrix wall.

Keywords: tissue engineering, cell transplantology, tissue engineering vascular implant, bioresorbable matrices, poly-lactide, electroforming

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