- Figueira T.R., Barros M.H, Camargo A.A., Castilho R.F., Ferreira J.C., Kowaltowski A.J., Sluse F.E., Souza-Pinto N.C., Vercesi A.E. 2013. Mitochondria as a source of reactive oxygen and nitrogen species: from molecular mechanisms to human health. Antiox. Redox Signal. 18 : 2029–2074.
- *Gaschler M.M., Stockwell B.R.* 2017. Lipid peroxidation in cell death. Biochem. Biophys. Res. Commun. 482 : 419–425.
- *Ghibelli L., Diederich M.* 2010. Multistep and multitask Bax activation. Mitochondrion. 10 : 604–613.
- Gusdon A.M., Fernandez-Bueno G.A., Wohlgemuth S., Fernandez J., Chen J., Mathews C.E. 2015. Respiration and substrate transport rates as well as reactive oxygen species production distinguish mitochondria from brain and liver. BMC Biochem. 17 p.

https://doi.org/10.1186/s12858-015-0051-8

- Habeeb A.F.S.A. 1972. Reaction of protein sulfhydryl groups with Ellman's reagent. Meth. Enzymol. 34 : 457–464.
- Halliwell B., Gutteridge M.C. 2015. Free radicals in biology and medicine. 5th edition. Oxford: Oxf. Univ. Press. 896 p.
- Handy D.E., Loscalzo J. 2012. Redox regulation of mitochondrial function. Antiox. Redox Signal. 16 : 1323–1367.

Lakowicz J.R. 2000. On spectral relaxation in proteins. Photochem. Photobiol. 72 : 421–437.

- *LoConte M., Carroll K.S., Jakob U.* 2012. The chemistry of thiol oxidation and detection. In: Oxidative stress and redox regulation. N.-Y.: Springer. 1–42.
- Lowry D.H., Rosembrough H.J., Farr A.L. 1951. Protein measurement with the Folin-phenol reagent. J. Biol. Chem. 193: 265–275.
- *Maroz A., Anderson R.F., Smith R.A., Murphy M.P.* 2009. Reactivity of ubiquinone and ubiquinol with superoxide and the hydroperoxyl radical: implications for *in vivo* antioxidant activity. Free Rad. Biol. Med. 46 : 105–109.
- *Mihaljević B., Katušin-Ražem B., Ražem D.* 1996. The reevaluation of the ferric thiocyanate assay for lipid hydroperoxides with special considerations of the mechanistic aspects of the response. Free Rad. Biol. Med. 21 : 53–63.
- Pacher P., Beckman J.S., Liaudet L. 2007. Nitric oxide and peroxynitrite in health and disease. Physiol. Rev. 87: 315–424.

- Panel M., Ghaleh B., Morin D. 2018. Mitochondria and aging: A role for the mitochondrial transition pore? Aging Cell. 17(4). https://doi.org/10.1111/acel.12793
- *Polderman K.H.* 2009. Mechanisms of action, physiological effects, and complications of hypothermia. Critical Care Med. 37 : 186–202.
- Schenkel L.C., Bakovic M. 2014. Formation and regulation of mitochondrial membranes. Int. J. Cell Biol. Article ID 709828. 13 p. https://doi.org/10.1155/2014/709828
- *Schrepfer E., Scorrano L.* 2016. Mitofusins, from mitochondria to metabolism. Mol. Cell. 61 : 683–694.
- *Shutt T., Geoffrion M., Milne R., McBride H.M.* 2012. The intracellular redox state is a core determinant of mitochondrial fusion. EMBO Rep. 13 : 909–915.
- Sies H., Berndt C., Jones D.P. 2017. Oxidative stress. Annu. Rev. Biochem. 86 : 715–748.
- *Søreide K.* 2014. Clinical and translational aspects of hypothermia in major trauma patients: from pathophysiology to prevention, prognosis and potential preservation. Injury. 45:647–654.
- Sun Z., Honar H., Sessler D.I., Dalton J.E., Yang D., Panjasawatwong K., Deroee A.F., Salmasi V., Saager L., Kurz A. 2015. Intraoperative core temperature patterns, transfusion requirement, and hospital duration in patients warmed with forced air. Anesthesiol. 122 : 276–285.
- *Venditti P., Rosa R.D., Meo S.D.* 2004. Effect of cold-induced hyperthyriodism on H_2O_2 production and susceptibility of stress conditions of rat liver mitochondria. Free Rad. Biol. Med. 36 : 348–358.
- Wong H.S., Dighe P.A., Mezera V., Monternier P.A., Brand M.D. 2017. Production of superoxide and hydrogen peroxide from specific mitochondrial sites under different bioenergetic conditions. J. Biol. Chem. 292 : 16804–16809.
- Zinchuk V.V., Hlutkin S.V. 2015. Blood oxygen transport and prooxidant-antioxidant balance in rats under hypothermia and rewarming combined with modification of L-arginine-NO pathway. Asian J. Pharmacy Nurs. Med. Sci. 3: 55–63.

INTENSITY OF FREE-RADICAL PROCESSES IN RAT LIVER MITOCHONDRIA AT MODERATE HYPOTHERMIA OF VARIOUS DURATION

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Artificial moderate hypothermia is widely used in clinical practice to protect organs from the effects of ischemia/reperfusion, injury and hypoxia. However, a decrease in the body temperature of homoiothermal animals induces oxidative stress, the severity of which may depend on the time of exposure to the "cold" factor. Since mitochondria play a key role in the generation of ROS, we studied the dependence of the intensity of FRP in mitochondria of the liver of rats on the duration of moderate (30°C) hypothermia. It turned out that short-term (30 min) hypothermia activates the processes of LP, while the concentration of lipid hydroperoxides, SchB and MDA significantly increases. Prolonging hypothermia to 1 hour reduces the content of many LP products, and at 3-hour hypothermia their normalization is observed. Short-term hypothermia and its prolongation to 1 hour is accompanied by oxidative destruction of mitochondrial proteins, which is reflected in a decrease in the content of sulfhydryl groups in them and an increase of carbonyl groups. At the same time, 3-hour hypothermia contributes to the normalization

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of the studied OMP markers. The dynamics of changes in the levels of sulfhydryl and carbonyl groups in mitochondrial matrix proteins is more pronounced in comparison with membrane proteins. The study of the spectral characteristics of membrane proteins of mitochondria showed a decrease in the intensity of their fluorescence in the initial stages of hypothermia. The main contribution to it is made by tryptophan residues localized at the periphery. The prolongation of hypothermia to 3 hours promote to restore the parameters of fluorescence to the level of control. The data obtained in the analysis of second derivatives of fluorescence spectra indicate certain changes in spatial configuration of membrane proteins.

Keywords: rats, hypothermia, liver, mitochondria, lipid peroxidation, oxidative modification of proteins

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